

ASCLEPIDACEAE

TRITERPENOIDS FROM THE ROOTS OF *HEMIDESMUS INDICUS*

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Key Word Index—*Hemidesmus indicus*; Asclepidaceae; lupeol octacosanoate; β -amyirin acetate.

Plant. *Hemidesmus indicus* R. Br. *Uses.* Medicinal.¹ *Previous work.* On roots,²⁻⁴ on leaves.⁵

Present work. The petrol extract of *H. indicus* roots on repeated chromatographic separation yielded, according to increasing order of polarity, hexatriacontane,⁶ lupeol octacosanoate (a new ester), β -amyirin acetate, lupeol acetate, α -amyirin, lupeol, β -amyirin and sitosterol. The new ester was found to have m.p. 81–82°, $[\alpha]_D +17.7^\circ$ (CHCl₃) and analyse for C₅₈H₁₀₄O₂. Strong IR absorptions at 1740 and 1175 cm⁻¹ was suggestive that it is an aliphatic ester, while absorptions at 1640 and 880 cm⁻¹ indicated a terminal double bond. On saponification with 6% ethanolic KOH it furnished a triterpene alcohol, C₃₀H₅₀O, m.p. 212–213°; $[\alpha]_D +30.4^\circ$ (CHCl₃) identical with lupeol (m.p., m.m.p., $[\alpha]_D$, IR; m.p., m.m.p., $[\alpha]_D$ and IR of acetate) and an acid, C₂₈H₅₆O₂, m.p. 82–84°. On esterification with CH₂N₂ the acid yielded a methyl ester, C₂₉H₅₈O₂, m.p. 60–62°. MS gave the molecular ion peak at *m/e* 438 and other prominent peaks at *m/e* 410, 396, 382, 354, etc. characteristic of a straight chain fatty acid methyl ester identical with methyl octacosanoate.⁷ Consequently, the new ester was characterized as lupeol octacosanoate.

EXPERIMENTAL

M.p.s are uncorrected. IR spectra were recorded as Nujol mulls, the NMR spectrum was taken at 60 MHz in CDCl₃ and TMS as internal standard and the MS was at 70 eV.

Extraction and chromatography. Dried roots of *H. indicus* (900 g) were exhaustively extracted with petrol. (60–80°) for 60 hr and the dark brown residue (40.2 g) obtained after removal of the solvent was separated into six distinct fractions, 1 (10.1 g), 2 (4.6 g), 3 (2.7 g), 4 (4.2 g), 5 (2.6 g), 6 (2 g) by chromatography over alumina (1 kg) and successive elution with petrol., petrol.–benzene (4:1), benzene and benzene–Et₂O mixture (1:1).

Fraction 1 on rechromatography over alumina (300 g) was separated into two fractions, Ia and Ib. Ia on crystallization from CHCl₃ yielded needles (200 mg), m.p. 74–75° identical with hexatriacontane⁶ (m.p., m.m.p., IR and co-TLC). Ib on crystallization from *n*-hexane yielded lupeol octacosanoate (3.3 g) m.p. 81–82°; $[\alpha]_D^{27} +17.7^\circ$ (CHCl₃); IR 1740, 1175 (ester carbonyl), 880, 1640 (terminal double bond), 715 cm⁻¹ (methylene chain): NMR δ 0.82 (s, 3H), 0.85 (s, 9H), 0.96 (s, 3H), 1.06 (s, 3H), 1.31 (broad s, 50H) 1.71 (broad s, 3H), 1.92–2.5 (m, 4H), 4.33–4.85 (m, 3H) (Found: C, 83.72; H, 12.82. C₅₈H₁₀₄O₂ requires: C, 83.65; H, 12.50%). Lupeol octacosanoate on saponification furnished lupeol, m.p. 212–213°; $[\alpha]_D +30.4^\circ$ (CHCl₃); IR 3250 (hydroxyl), 880 cm⁻¹ (=CH₂) and octacosanoic acid, m.p. 82–84°; IR 1705 (acid carbonyl),

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³ A. T. DUTTA, S. GHOSH and R. N. CHOPRA, *Arch. Pharm.* **276**, 333 (1938).

⁴ P. B. RAMAMURTI and T. R. SESHADRI, *Proc. Indian Acad. Sci.* **13A**, 399 (1941).

⁵ S. S. SUBRAMANIAN and A. G. R. NAIR, *Phytochem.* **7**, 1703 (1968).

⁶ I. HEILBRON, *Dictionary of Organic Compounds*, Vol. II, p. 677, Oxford University Press, Oxford (1953).

⁷ C. F. KREWSON, *J. Am. Chem. Soc.* **73**, 1365 (1951).

3300 (H-bonded hydroxyl), 718, 728 cm^{-1} (methylene chain) (Found C, 79.28; H, 13.26. Calc. for $\text{C}_{28}\text{H}_{56}\text{O}_2$: C, 79.24; H, 13.20%). The acid on esterification with CH_3N_2 yielded its methyl ester; IR 1740 (ester carbonyl), 719, 729 cm^{-1} (methylene chain) (Found: C, 79.48; H, 13.26. Calc. for $\text{C}_{29}\text{H}_{58}\text{O}_2$: C, 79.45; H, 13.24%).

Fraction 2 (4.6 g) was separated into fractions 2a and 2b by rechromatography over AgNO_3 impregnated silica gel. Fraction 2a on crystallization from CHCl_3 -MeOH yielded β -amyrin acetate in needles (700 mg), m.p. and m.m.p. 234–236°; $[\alpha]_D^{27} +79^\circ$ (CHCl_3). Fraction 2b on crystallization from CHCl_3 -MeOH afforded lupeol acetate (2.8 g), m.p. and m.m.p. 216–218°; $[\alpha]_D^{27} +42^\circ$ (CHCl_3).

Fraction 3 (2.7 g) on rechromatography on AgNO_3 impregnated silica gel yielded α -amyrin and lupeol (characterized by m.p., m.m.p., co-TLC, IR). Fraction 4 on rechromatography over alumina yielded β -amyrin and sitosterol (characterized by m.p., m.m.p., co-TLC, IR). Fraction 5 on further purification by chromatography over alumina afforded more sitosterol (characterized by m.p., m.m.p., co-TLC, IR).

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BERBERIDACEAE

ANTHOCYANINS IN FRUITS OF *BERBERIS BUXIFOLIA*

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Key Word Index—*Berberis buxifolia*; Berberidaceae; petunidin; peonidin; malvidin and delphinidin glycosides.

The genus *Berberis* (Berberidaceae) has not been fully investigated although nearly 150 species are distributed all over the world. Recently Mamaev and Semkina^{1,2} identified the anthocyanins of *Berberis thunbergii* and *B. vulgaris* (common barberry) and observed a maximum pigment concentration in spring. The leaves of the purple-leaf and green-leaf forms of barberry contained five anthocyanin pigments, the main ones being 3-monoglycosides of peonidin, cyanidin and delphinidin. No reports on isolation of anthocyanins from *Berberis* fruits are known.

The present work describes the identification of ten anthocyanins isolated from *Berberis buxifolia* Lam. fruits, which is indigenous to Argentina and south of Chile. Chromatography of the crude extract yielded six coloured bands, in amounts decreasing in the order IV > III \geq II > I > V > VI, the latter band being feint. Each band was rechromatographed in 15% HOAc to give complete purification. R_f s are shown in Table 1. Only pigments IIIa, IIIb, IVb, IVc, Va and Vb changed to blue with 1% ethanolic $\text{Pb}(\text{OAc})_2$.³

The visible and UV spectra of pigments IVa and Va were characteristic of 3,5-diglycosides and only differed by substitution in the B-ring.⁴ The other spectra corresponded to 3-glyco-

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